Survey and Diagnostics

Subcommittee
Update
Q TAC Meeting
St Louis, MO
April 3, 2006



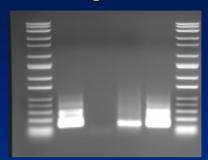
Cindy McKenzie and Frank Byrne

Monitoring guidelines

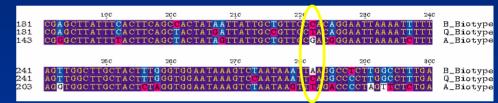
- Grower-submitted vs State samples:
 - Keeping growers anonymous ensured successful implementation of this component of the survey.
 - report at state level.
- State samples:
 - A-list 30 sample locations, B-list 10 sample locations.
 - Min # insects per sample = 10.
 - Proportion of samples obtained from GH = 60-80%, other = 20-40%.
 - report at county level.

Detection Techniques

- Based on Genetic differences
- Electrophoresis
- PCR
- Gene sequencing
 - COI
 - Microsatellites







- Labs Conducting Biotype Analysis
 - Judy Brown, U AZ COI
 - Frank Byrne, U CA Esterase
 - Cindy McKenzie/Bob Shatters, ARS
 COI and Microsatellites



Biotype B

Biotype Q Africa B-related Asia I

Asia II Biotype A

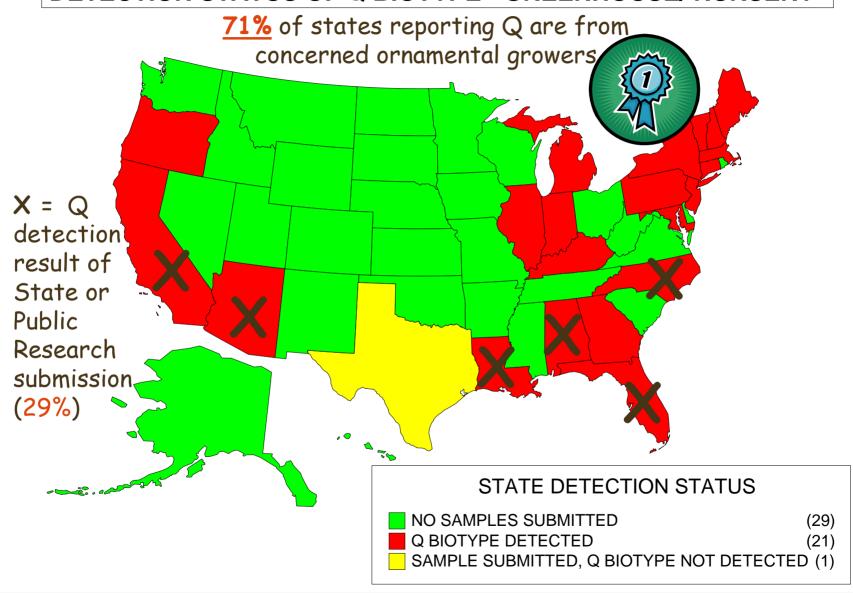
Sub-Saharan Africa

Africa

North American "Q" Biotype Detections



DETECTION STATUS OF Q BIOTYPE - GREENHOUSE/ NURSERY



Conclusions

- More than one positive Q sample in some states 21 states and counting.
- States identified as positive not overrun with Q; all populations were managed.
- Microsatellite data confirms multiple introductions.
- No positive ids in anything other than ornamentals and herbs; unofficial report on tomato transplants from retail outlet in AZ.
- Ornamentals includes a lot of hosts, not just poinsettias.

Survey and Diagnostics Top Research Priorities "Grower Friendly Diagnostics"

Short Term (12 months)

- 1. Training more Labs doing Diagnostics.
- 2. In-Depth Microsatellite Study with more locations and more alleles.

Long Term (5 year plan)

- 1. Correlate markers w/insecticide profiles.
- 2. Identification of B & Q specific protein and genetic markers (ELISA).

Microsatellite Study

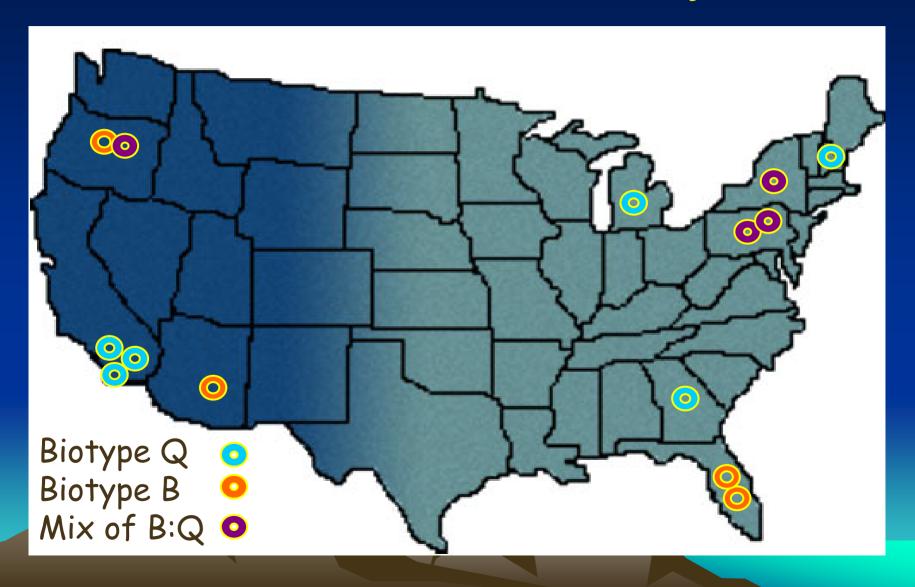
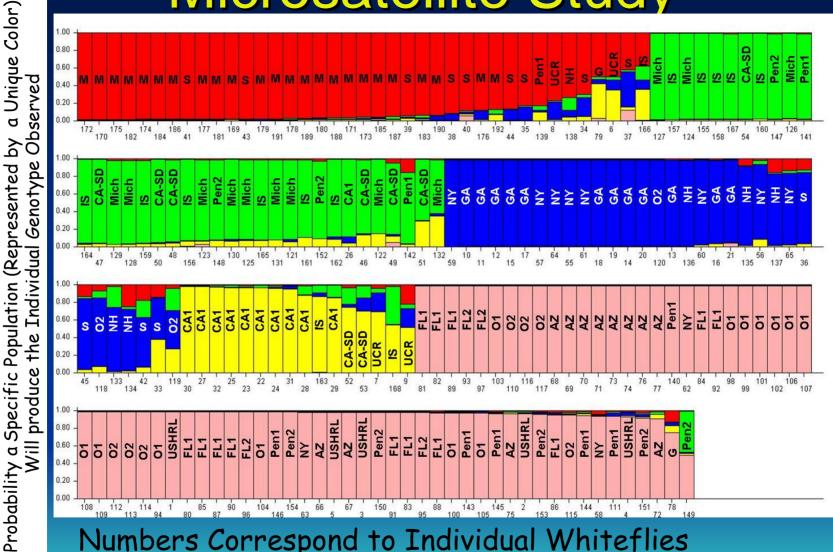


TABLE 2. Description of *Bemisia tabaci* populations used in this study.

Population	Abreviatio	Location	COI Biotype	Individual
Number	n		Determination	Numbers
1	USHRL	USHRL Lab Colony	В	1-5
2	UCR	UC Riverside Quarantine	Q	6-9
3	GA	Georgia	Q	10-21
4	CA1	California1	Q	22-32
5	5	Spain	Q	33-45
6	CA-SD	California-SD	Q	46-54
7	NY	New York	B and Q	55-65
8	ΑZ	Arizona	В	66-77
9	G	Guatemala	222	78-79
10	FL1	Florida1	В	80-92
11	FL2	Florida2	В	93-97
12	<i>O</i> 1	Oregon1	В	98-109
13	02	Oregon2	B and Q	110-120
14	Mich	Michigan	Q	121-132
15	NH	New Hampshire	Q	133-138
16	Pen1	Pennsylvania1	B and Q	139-146
17	Pen2	Pensylvania2	B and Q	147-154
18	I5	Israel1	Q	155-160
19	IS	Isreal2	Q	161-168
20	M	Morocco1	Q	169-180
21	M	Morocco2	Q	181-192

Microsatellite Study



Numbers Correspond to Individual Whiteflies

Microsatellite Study (Individuals Grouped by Population)

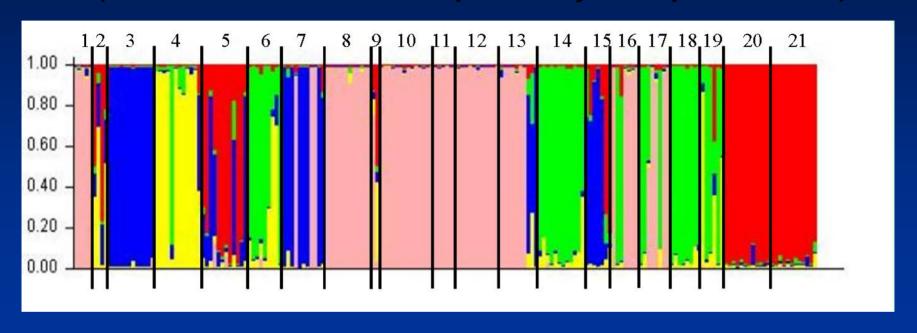


Table 3. Description of Q Biotype Subgroups

Q Subgroup	Old World Representation	U.S. Representation
Subgroup 1 (Red)	Morocco/Spain	Pennsylvanina, New Hampshire, UCR-quarantine colony
Subgoup 2 (Green)	Israel	California, Michigan, Pennsylvania,
Subgroup 3 (Blue)	Spain	Georgia, New Hampshire, New York, Oregon
Subgroup 4 (Yellow)	Israel	California, UCR-quarantine colony (???)

Summary

- All Q biotype whiteflies analyzed to date (from the U.S., Spain, Morocco, and Israel) can be subdivided into four separate subgroups, all of which are in the United States.
- The Four U.S. Q Biotype Subtypes Suggest Multiple Introductions of Biotype Q into the U.S.
- The Q biotype has much greater microsatellite diversity than observed for the B biotype in the U.S. The genetic diversity of the Q biotype is similar to that reported for the indigenous Asia-Pacific genotypes (De Barro, 2005).
- · Our data show that microsatellite genotyping is powerful enough to distinguish among subtypes of the Q biotype. Future work coordinating the microsatellite genotyping with insecticide resistance profiles will be conducted to determine if this genotyping method can be used as a predictor of insecticide resistance profiles.