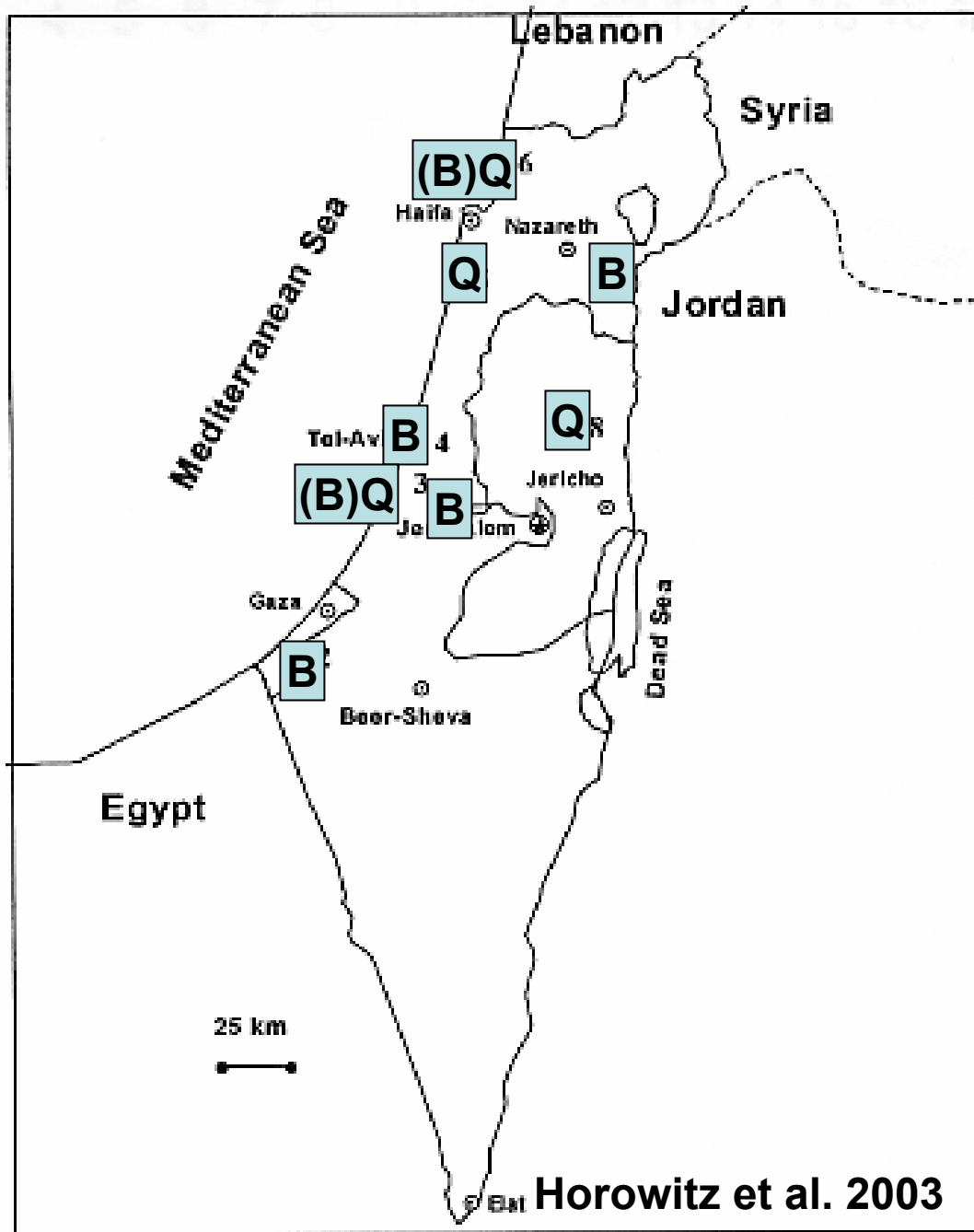


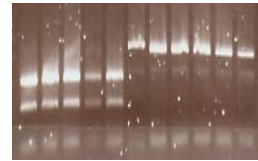
# Evolution of insecticide resistance in the B and Q sympatric biotypes of *Bemisia tabaci* - single or multiple origins?

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**Faculty of Agriculture, The Hebrew University of Jerusalem**



Q Biotype    B Biotype



**Are the B and Q biotypes of  
*Bemisia tabaci* genetically  
isolated?**

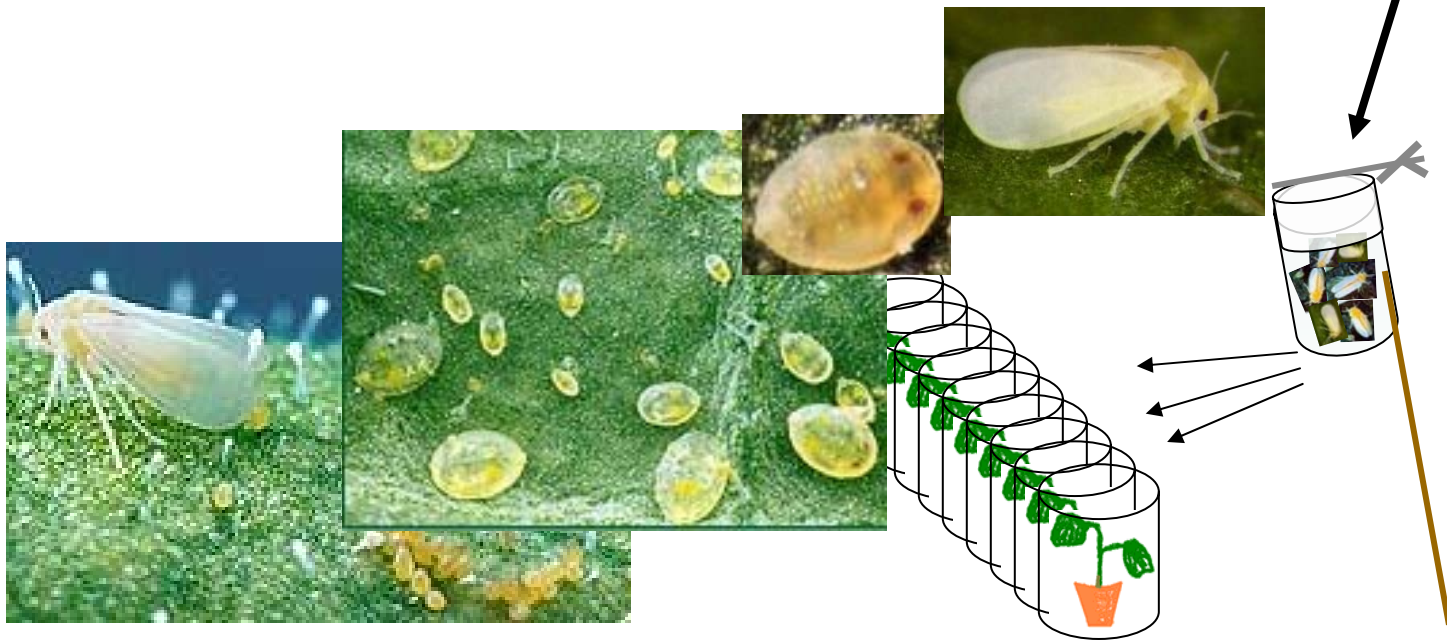
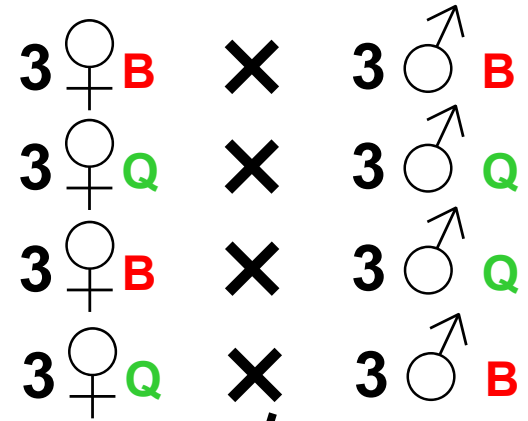
# Strains used in the study

<b>Strain</b>	<b>Biotype</b>	<b>Year</b>	<b>Country</b>	<b>Host</b>
<b>B ref</b>	<b>B</b>	<b>1987</b>	<b>Israel</b>	<b>Cotton</b>
<b>Ashalim</b>	<b>B</b>	<b>2003</b>	<b>Israel</b>	<b>Melons</b>
<b>PyriS</b>	<b>Q</b>	<b>1991</b>	<b>Israel</b>	<b>Roses</b>
<b>Ayalon</b>	<b>Q</b>	<b>2002</b>	<b>Israel</b>	<b>Cotton</b>

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<b>Ayalon</b>	<b>Q</b>	<b>2002</b>	<b>Israel</b>	<b>Cotton</b>

# Cross design for inter and intra biotype mating

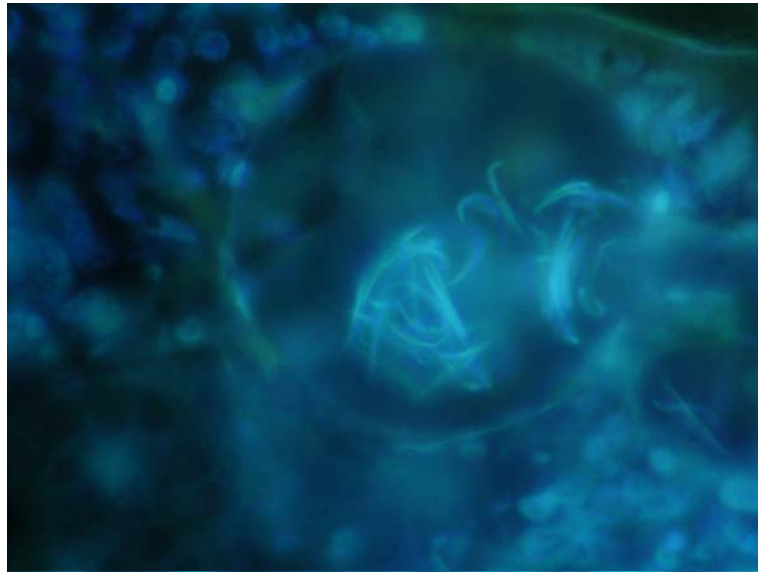


1. While males and females progeny are produced within each biotype, no female progeny are produced by crosses between biotypes

**B Ashalim X Q Ayalon**

	 <b>B</b>	 <b>Q</b>
 <b>Q</b>	Total ♀ = 0 Total ♂ = 78	Total ♀ = 46 Total ♂ = 90
 <b>B</b>	Total ♀ = 26 Total ♂ = 35	Total ♀ = 0 Total ♂ = 78

## 2. No sperm is transferred between biotypes – Pre-zygotic barrier



**BXB**

**QXQ**

**BXQ or Virgin  
Females**





### 3. Disturbed courtship and mating behavior between the B and Q biotypes



5. Wing movement



6. Copulatory organ contact.



1. Female contact and male orientation.



4. Body pushing



3. Abdominal movement



2. Antennal drumming.

## 4. The possible involvement of symbionts is unlikely

Strain/symbionts	<i>Wolbachia</i> sp.	<i>Rickettsia</i> sp.	<i>Hamiltonella</i> <i>defensa</i>
<b>B Ashalim</b>	---	-- / +	+++
<b>B ref</b>	---	+++	+++
<b>Q Ayalon</b>	---	-- / +	---
<b>Q PyriS</b>	---	+++	---

## 5. Unfertile female progeny are produced by crosses between biotypes collected ~20 years ago

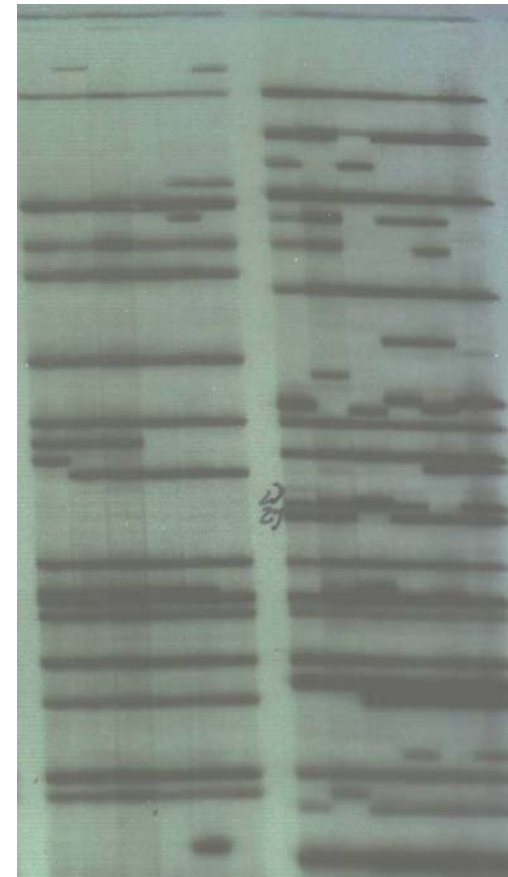
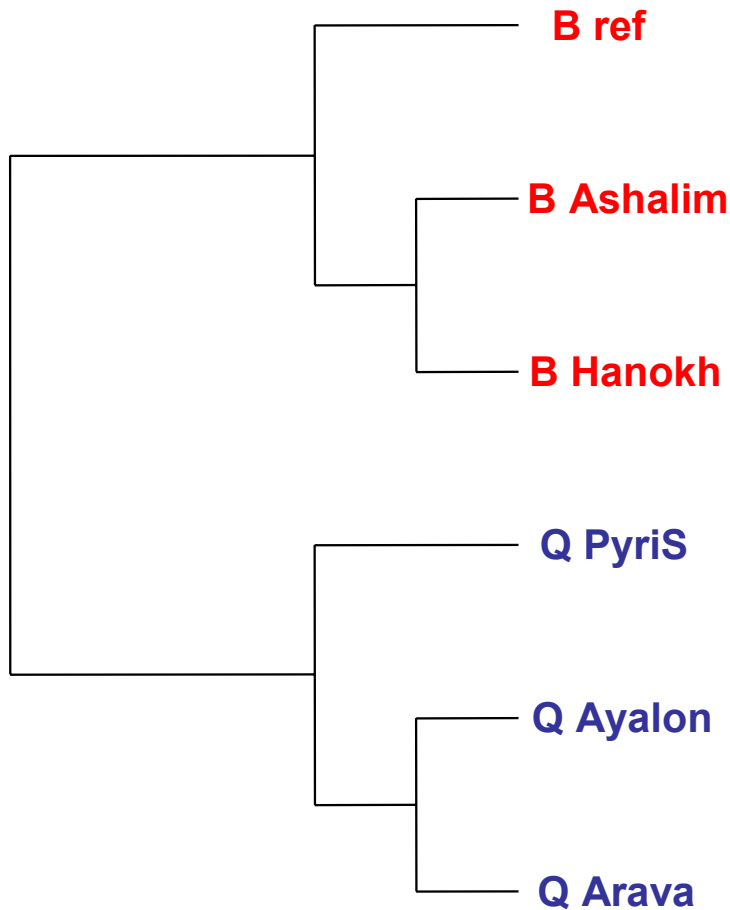
**B ref X Q PyriS**

	<u>♂</u> B	<u>♂</u> Q
<u>♀</u> Q	Total ♀=0 Total ♂= 136	Total ♀=29 Total ♂=18
<u>♀</u> B	Total ♀=77 Total ♂=102	Total ♀= <b>2BQ</b> Total ♂= 72

# 6. AFLP analysis of molecular variance between populations of the B and Q biotypes shows significant reduction in gene flow

Source of variation	d.f.	Sum of squares	Variance components	Percentage of variation
Between biotypes	1	535.333	33.87111 Va	67.84
Among populations within biotypes	4	109.067	2.80333 Vb	5.62
Within populations	24	318.000	13.25000 Vc	26.54
Total	29	962.400	49.92444	

	1	2	3	4	5	6
1 B Ashalim	-	0.07653	0.13265	0.48980	0.53571	0.52551
2 B Hanokh	15	-	0.09694	0.48469	0.52041	0.51020
3 B ref	26	19	-	0.46939	0.48469	0.48469
4 Q PyriS	96	95	92	-	0.13776	0.15816
5 Q Ayalon	105	102	95	27	-	0.16327
6 Q Arava	103	100	95	31	32	-

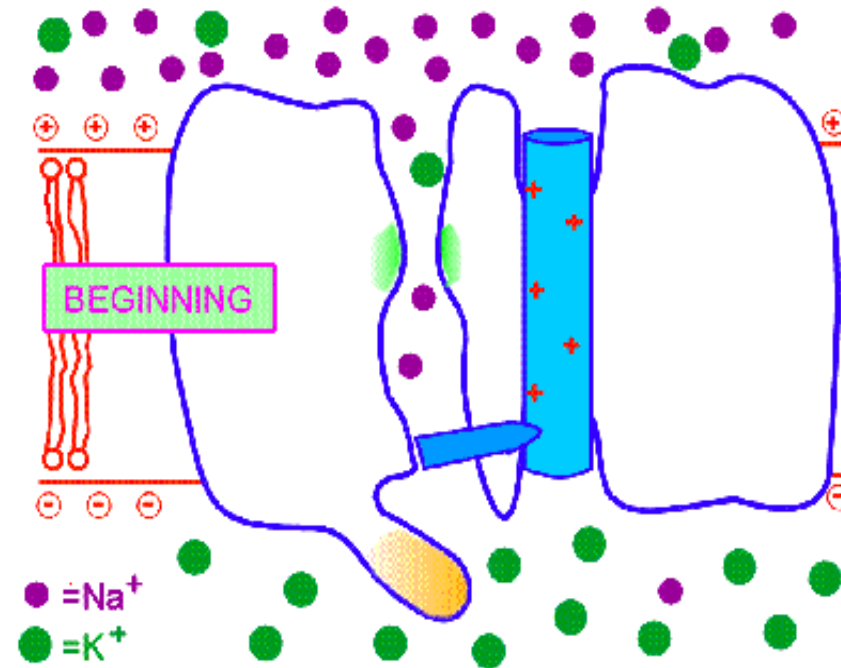
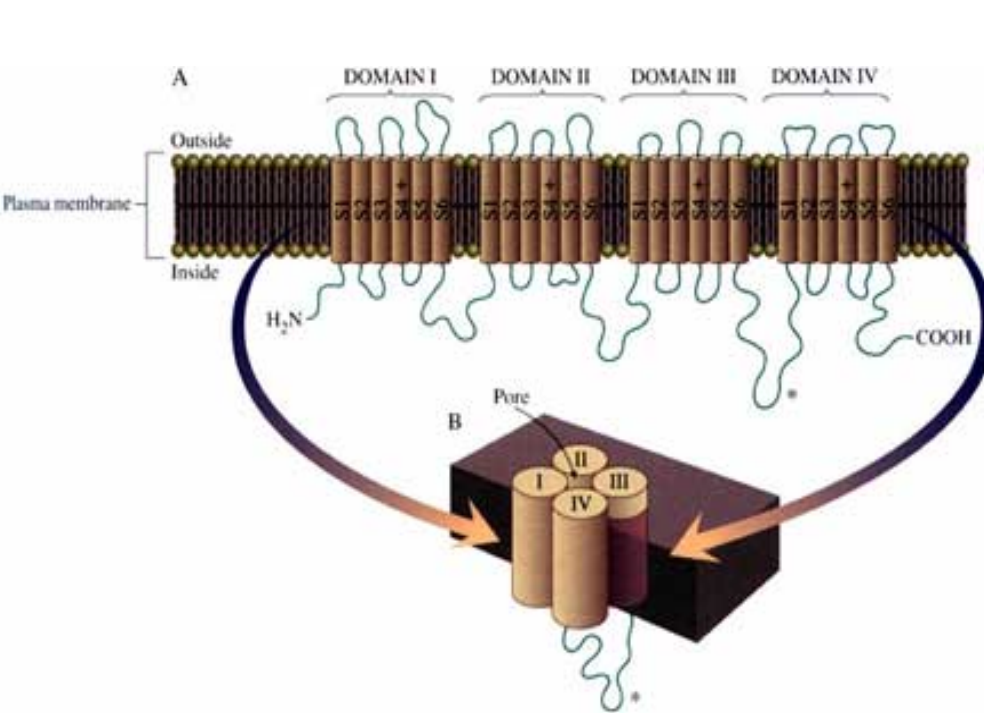


# Scientific Questions

- **Did similar insecticide selection in the B and Q biotypes result in similar genetic changes**
- **Did the genetic changes arise once in the common ancestor**
- **Arise once in one biotype and was subsequently transferred to the other biotype**
- **Arise independently in each biotype (parallel evolution)**

**Multiple Origins of Pyrethroid  
Resistance in Sympatric Biotypes  
of *Bemisia tabaci***

# The effect of pyrethroids on the *para*-type voltage gated sodium channel



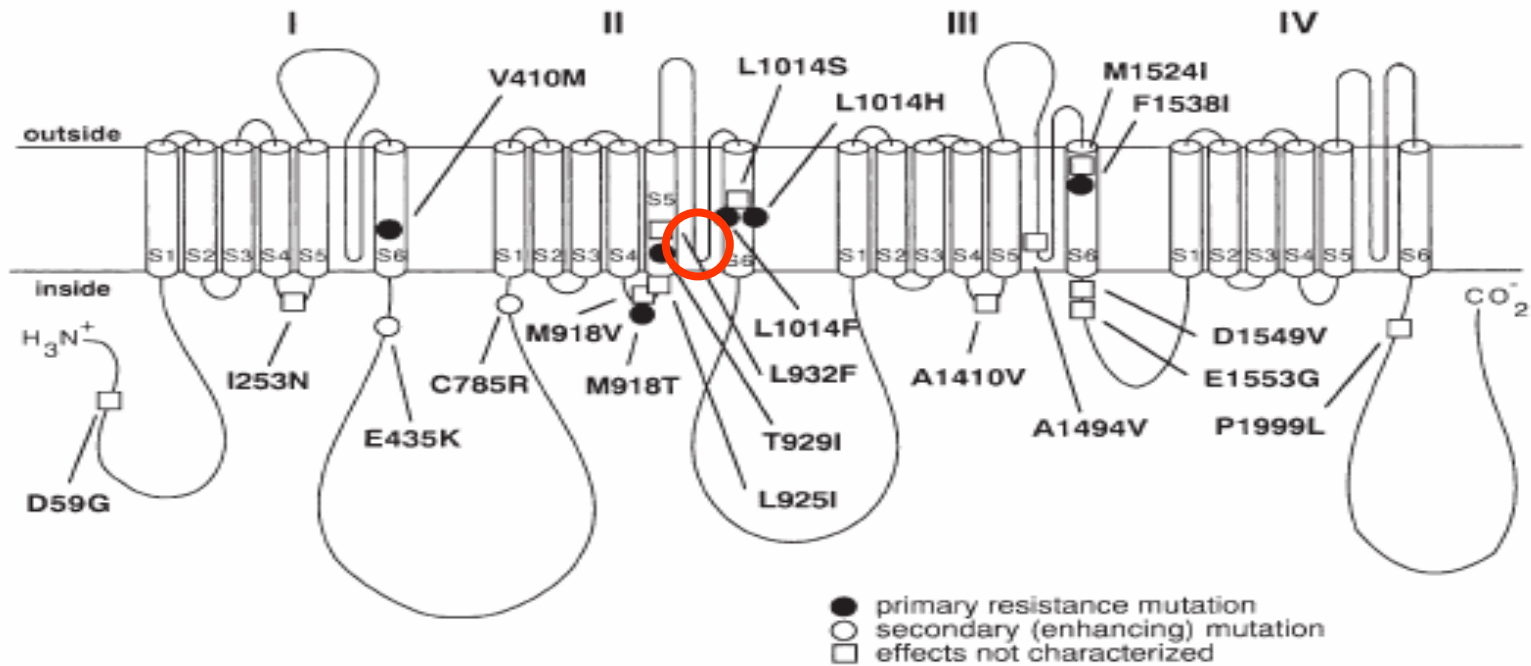
Control

After pyrethroid treatment



# Resistance to pyrethroids caused by mutations in the *para*-type voltage gated sodium channel

Resistance mechanism: lower sensitivity of the target site =  
Knockdown Resistance - kdr



Leu925Ile  
Met918 Thr929Val

<i>s-B</i>	GCCAAATCCTGGCCAACCTTGAATCTGTTGATTCAATCAATGGGCCGAACAGTTGGGGCC <b>TTA</b> GGAAATTG <b>ACT</b> TTTGT	80
<i>r1-B</i>	-----	80
<i>s-Q</i>	-----	80
<i>r1-Q</i>	-----	80
<i>r2-Q</i>	-----	80

<i>s-B</i>	TTTGTGATCATTATTTTCATTTTGTCTGTGATGGGAATGCAACTATTCGGGAAGAATTATACAGACAATGTTGATCGCT	160
<i>r1-B</i>	-----	160
<i>s-Q</i>	-----	160
<i>r1-Q</i>	-----	160
<i>r2-Q</i>	-----	160

Intron 1 (~730 bp)

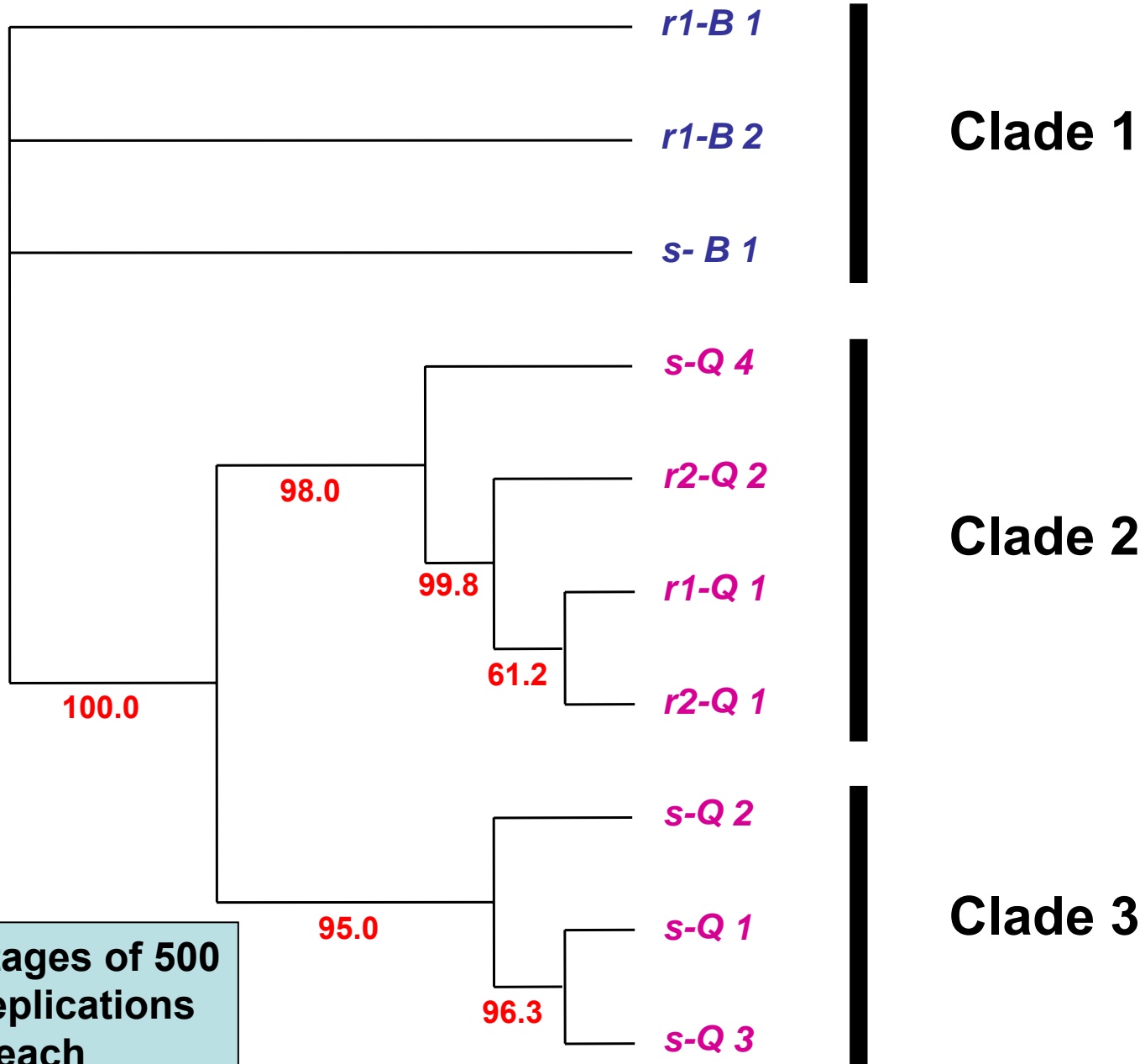
<i>s-B</i>	TTCTGGCGGAGAACTACCTCGGTGGAATTTACTGACTTCATGCACCTCATTTCATGATCGTTTTTCGAGTCTCTGCGGA	240
<i>r1-B</i>	-----	240
<i>s-Q</i>	-----	240
<i>r1-Q</i>	-----	240
<i>r2-Q</i>	-----	240

<i>s-B</i>	GAATGGATTGAGTCCATGTGGGACTGTATGCATGTTGGTGATGTGTCCTGTATTCCCTTTTTTTTTTAGCCACTGTCGTTAT	320
<i>r1-B</i>	-----	320
<i>s-Q</i>	-----	320
<i>r1-Q</i>	-----	320
<i>r2-Q</i>	-----	320

Leu1014

<i>s-B</i>	CGGTACCTTGTAGTTTAAATCTTTCTTAGCGTTGTTGCTGAGTAATTT	371
<i>r1-B</i>	-----	371
<i>s-Q</i>	-----	371
<i>r1-Q</i>	-----	371
<i>r2-Q</i>	-----	371

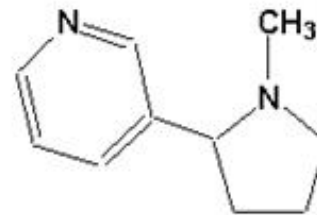
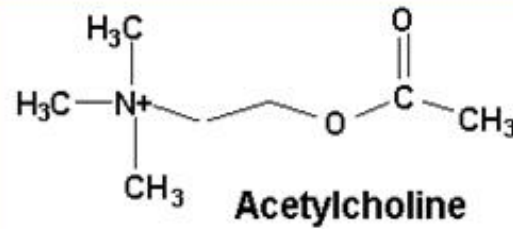
Intron 2



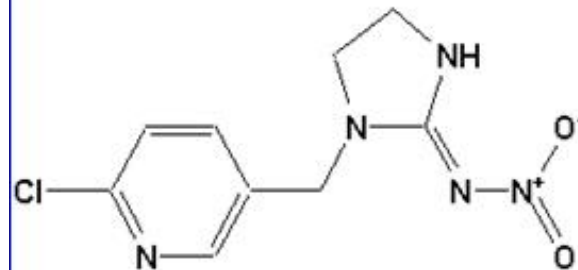
The percentages of 500 bootstrap replications supporting each branch are shown

**A Possibility for a Single Origin  
of Cytochrome P450  
Dependent Neonicotinoids  
Resistance in the B and Q  
biotypes of *Bemisia tabaci***

# Neonicotinoids - Nicotinic Acetylcholine receptor agonists

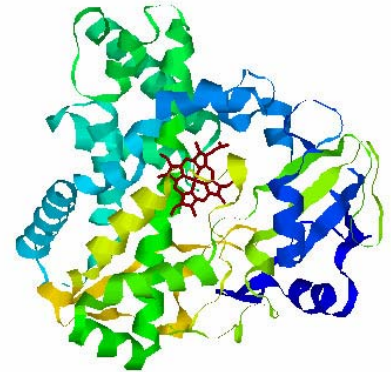
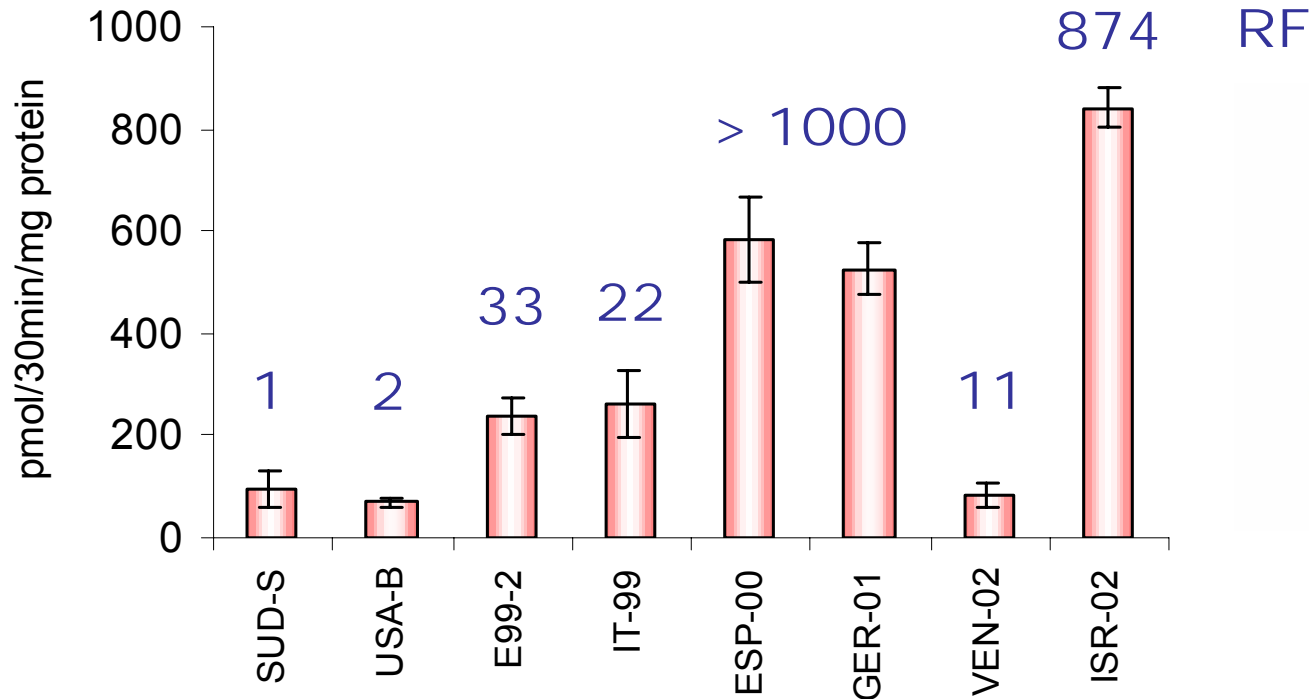
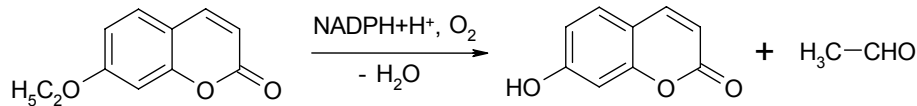


**Nicotine (55 mg/kg)**



**Imidacloprid (424-475 mg/kg)**

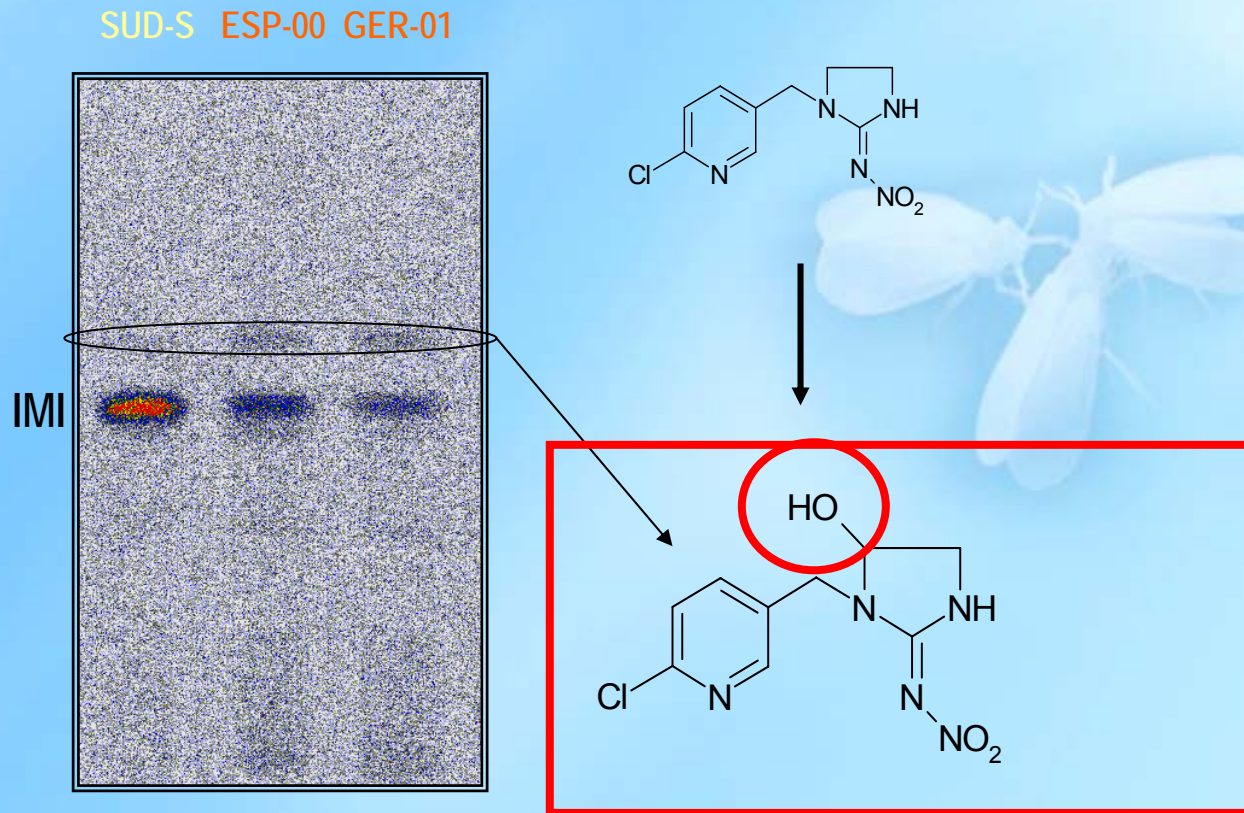
# P450s Activity Confer Neonicotinoid Resistance in *Bemisia tabaci*



Rauch & Nauen (2003) Arch. Insect Biochem. Physiol. 54

7-Ethoxycoumarin-O-deethylase activity is a biochemical marker linked to neonicotinoid resistance in *Bemisia tabaci*

# P450s Can Metabolize Imidacloprid to Five-Hydroxy-Imidacloprid



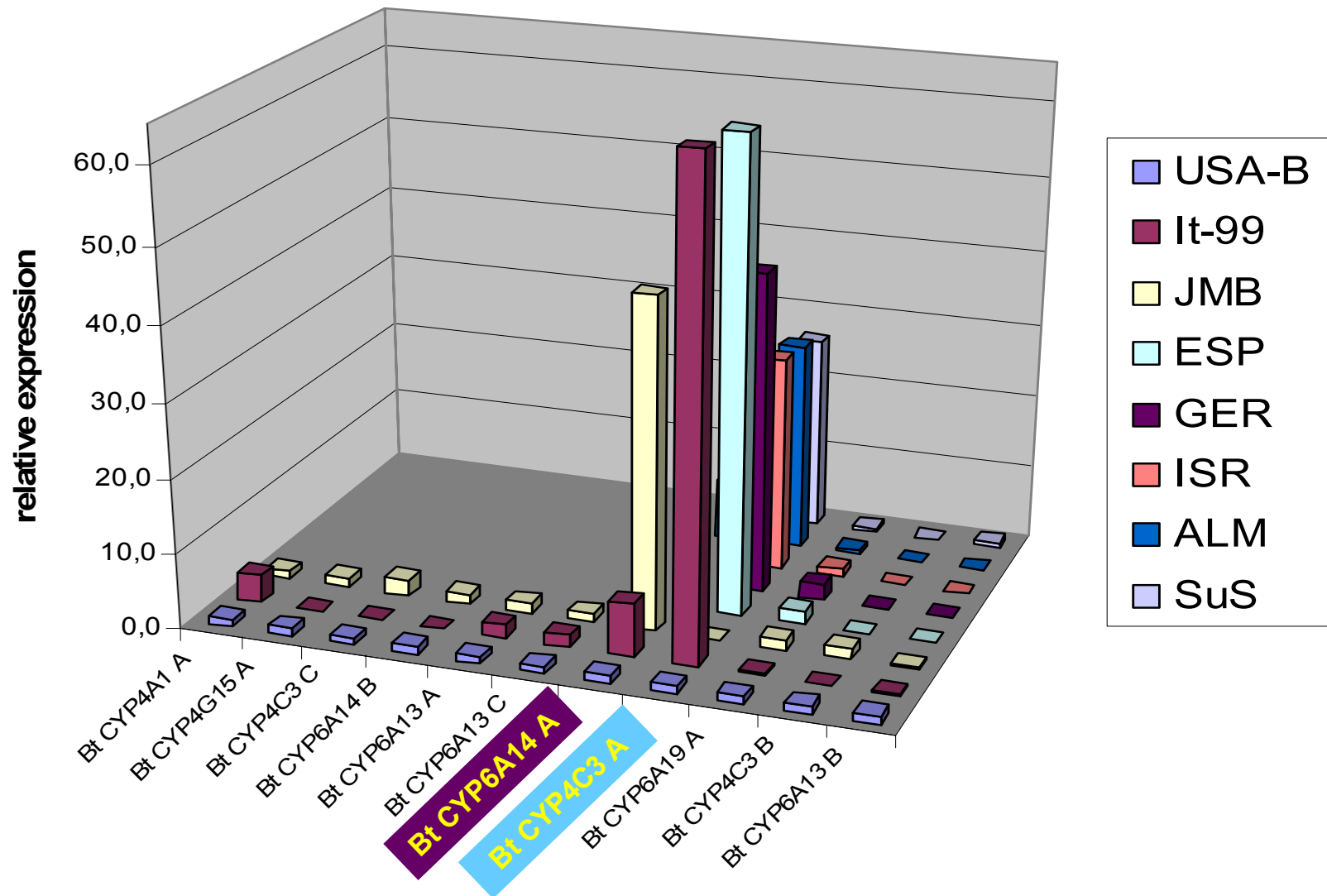
# Neonicotinoids Resistance is Found in Both the B and Q Biotypes

Strain	Year of collection	Origin	Biotype	Neonicotinoid-resistance	Resistance factor *
SUD-S	1978	Sudan	-	No	Reference
E99-2	1999	Spain	Q	Moderately	33
IT-99	1999	Italy	Q	Moderately	22
ESP-00	2000	Spain	Q	High	>1000
GER1-01	2001	Germany	Q	High	>1000
USA-B	1994	California	B	No	2
VEN-02	2002	Venezuela	B	Moderately	11
JMB	2003	Brazil	B	High	874 <sup>**</sup>

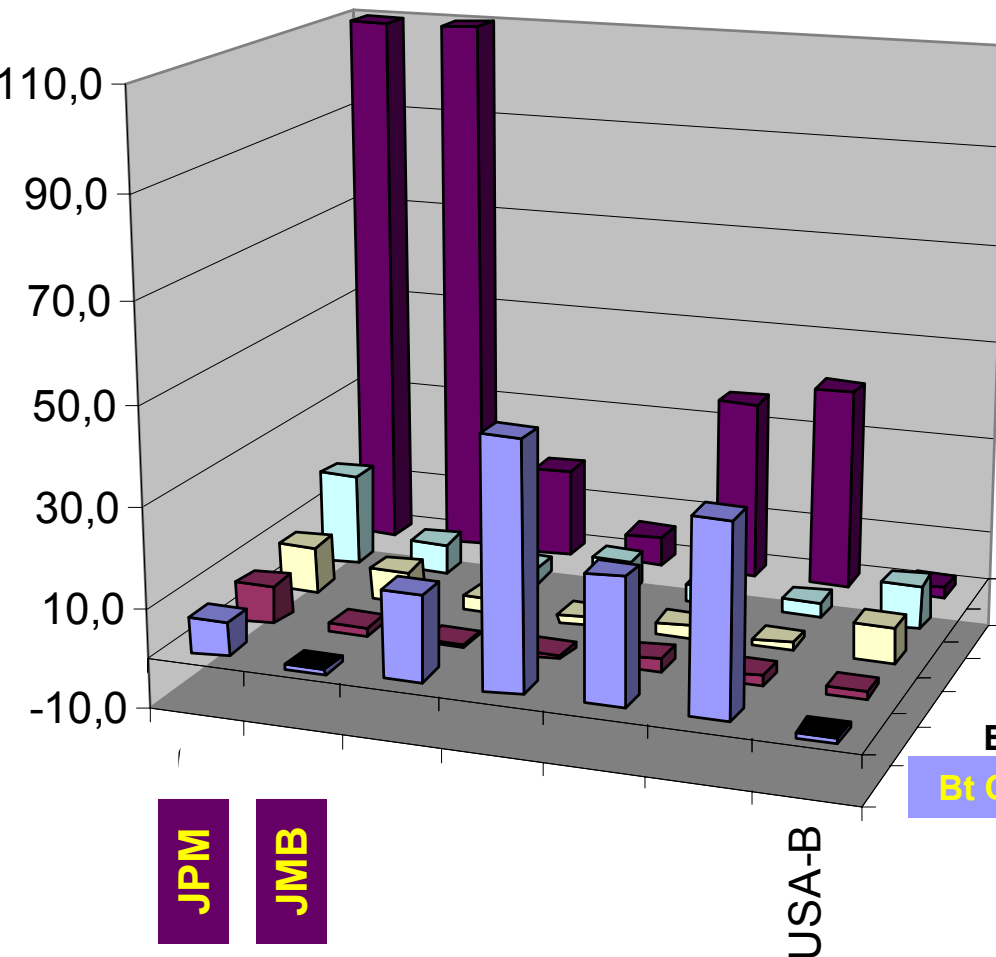
\* Leaf-dip bioassay with imidacloprid (72h)



# Real-time PCR-analysis of the 11 p450 genes showed that two of them are over-expressed in resistant strains



# Only one gene is correlated with resistance in both the B and Q biotypes



Strain	RF*	Comments
<b>JPB</b>	<b>+++</b>	<b>sys</b>
<b>JMB</b>	<b>+++</b>	<b>sys</b>
ALM	++	spray
IT	++	spray
GER	+++	leaf dip (2001)
ESP	+++	spray
<b>USA-B</b>	<b>No</b>	<b>leaf dip</b>
RF11-100	++	
RF>100	+++	

**Bt CYP6A14 A**

Bt CYP6A13 B

Bt CYP4C3 B

Bt CYP6A14 A

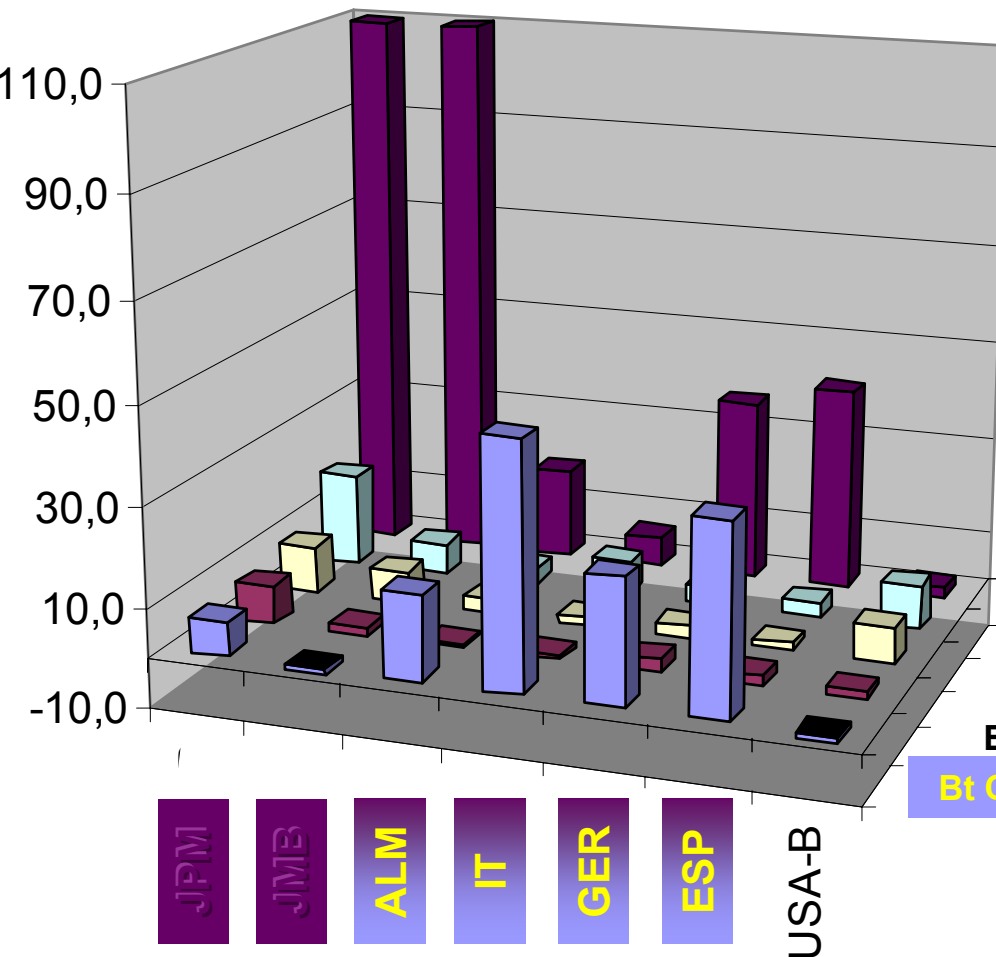
**Bt CYP4C3 A**

**JPM**

**JMB**

USA-B

# Only one gene is correlated with resistance in both the B and Q biotypes



Strain	RF*	Comments
JPB	+++	sys
JMB	+++	sys
ALM	++	spray
IT	++	spray
GER	+++	leaf dip (2001)
ESP	+++	spray
USA-B	No	leaf dip
RF11-100	++	
RF>100	+++	

Bt CYP6A14 A

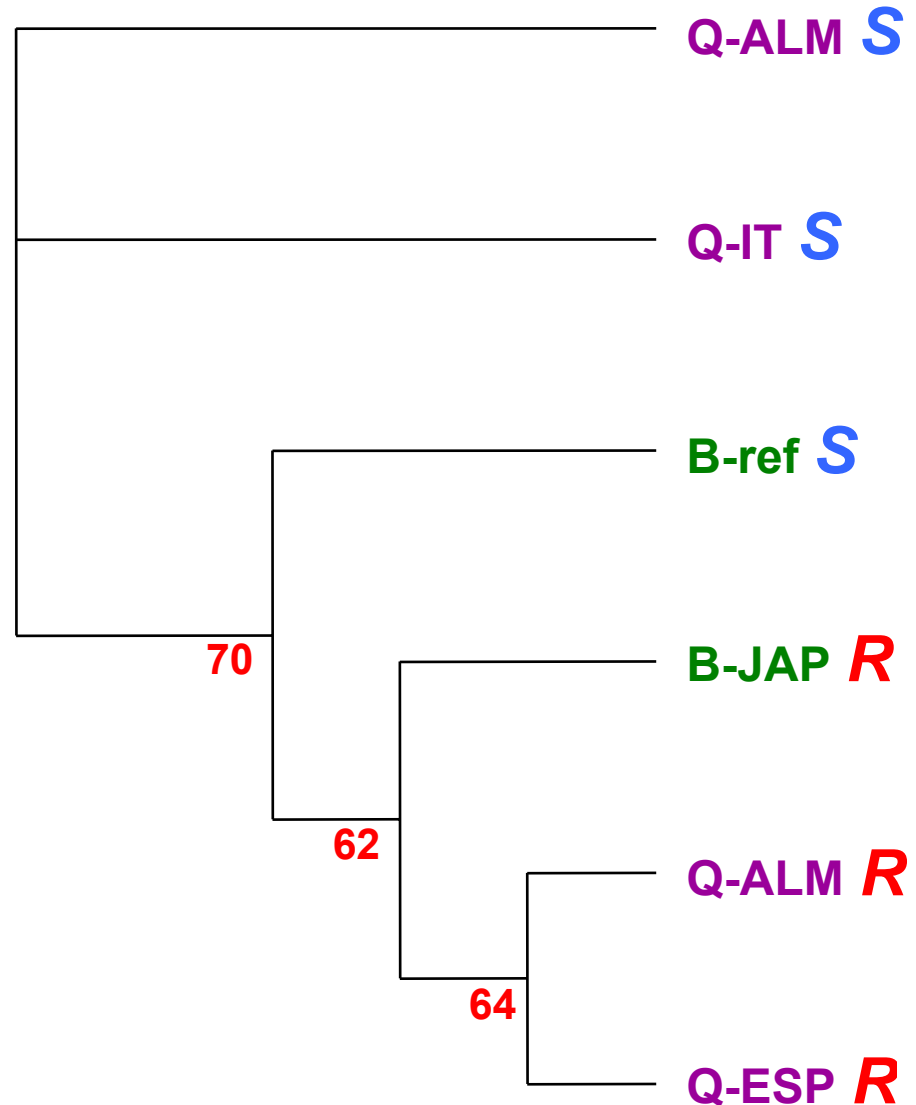
Bt CYP6A13 B

Bt CYP4C3 B

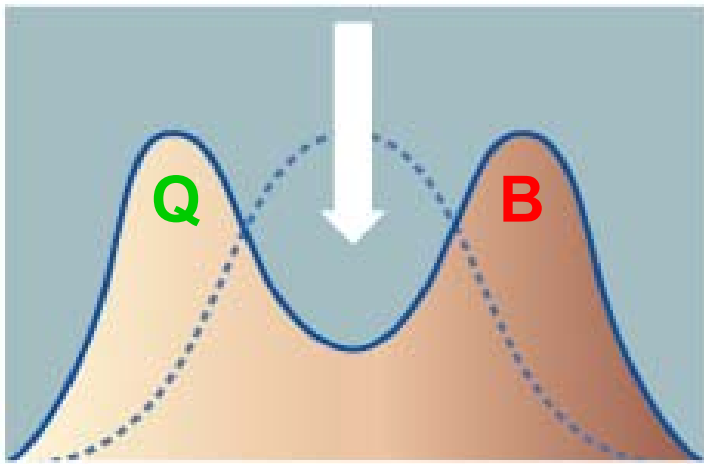
Bt CYP6A14 A

Bt CYP4C3 A

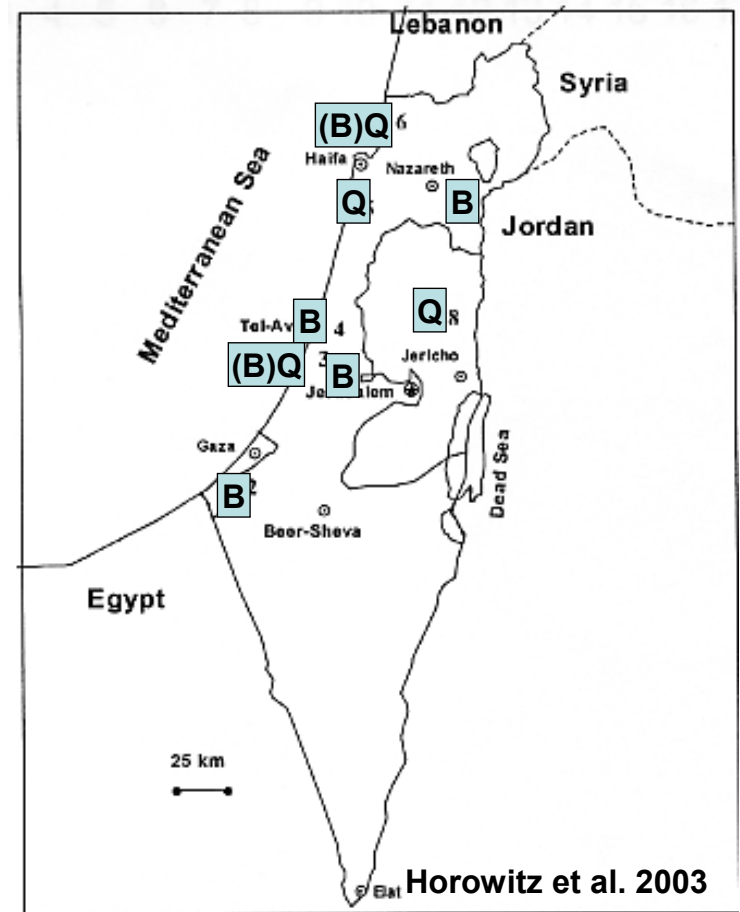
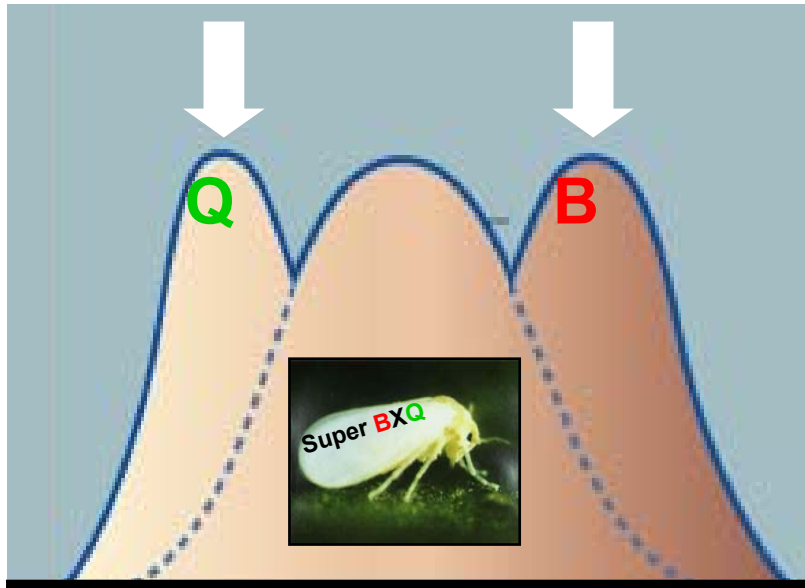
# Unrooted maximum likelihood tree of *Bemisia tabaci*, produced from the first 0.3 kb intron sequence of the Bt CYP6A14 A gene



The percentages of 500 bootstrap replications supporting each branch are shown



## Allopatric speciation with secondary contact



## Collaborators:

Juergen Benting	Bayer CropScience
Ralf Nauen	Bayer CropScience
Rami Horowitz	The Israeli Agricultural Research Organization

## Students in the lab:

Iris Karunker	P450s and neonicotinoid resistance
Navit Lahav	Reproductive isolation between biotypes
Michal Alon	Evolution of resistance to pyrethroids
Moshe Elbaz	Mating behaviour
Dina Berlovitch	Mating behaviour